



## Endo Agar

M029

### Intended Use

Endo Agar is a selective medium recommended for confirmation of the presumptive test for members of the coliform group from clinical and non-clinical samples.

### Composition\*\*

| Ingredients           | Gms / Litre |
|-----------------------|-------------|
| Peptone               | 10.000      |
| Lactose               | 10.000      |
| Dipotassium phosphate | 3.500       |
| Sodium sulphite       | 2.500       |
| Basic fuchsin         | 0.500       |
| Agar                  | 15.000      |
| Final pH ( at 25°C)   | 7.5±0.2     |

\*\*Formula adjusted, standardized to suit performance parameters

### Directions

Suspend 41.5 grams in 1000 ml purified/distilled water. Heat to boiling to dissolve the medium completely. Sterilize by autoclaving at 15 lbs pressure (121°C) for 15 minutes. Mix well before pouring into sterile Petri plates. If the solidified culture medium is somewhat too red, then to remove the colour add a few drops (max. 1 ml/litre) of a freshly prepared 10% Sodium sulphite solution and boil.

### Principle And Interpretation

Endo Agar was developed by Endo to differentiate gram-negative bacteria on the basis of lactose fermentation, while inhibiting gram-positive bacteria (2). Inhibition of the later was achieved without the use of bile salts as was traditionally used. Endo was successful in inhibiting gram-positive bacteria on his medium by the incorporation of sodium sulphite and basic fuchsin. The resulting Endo Agar, also known as Fuchsin Sulphite and Infusion Agar, was used to isolate the typhoid bacilli. Many modifications of this media have been done over the years. Endo Agar is recommended by APHA as an important medium in the microbiological examination of water and wastewater, dairy products and foods (1,5,6). Endo Agar is used to confirm the detection and enumeration of coliform bacteria following presumptive test of drinking water. It is also used for the detection and isolation of coliforms and faecal coliforms from milk, dairy products and food.

The medium contains peptone which provide nitrogen, carbon, vitamins and minerals required for bacterial growth. Sodium sulphite and basic fuchsin make this medium selective by suppressing gram-positive organisms. Coliforms produce pink colonies on fermentation of lactose while lactose non-fermenters produce colourless colonies on the medium.

With *Escherichia coli*, this reaction is very pronounced as the fuchsin crystallizes, exhibiting a permanent greenish metallic luster (fuchsin luster) to the colonies. Medium should be stored away from light to avoid photo-oxidation.

### Type of specimen

Clinical samples - faeces ; Food and dairy samples; Water samples

### Specimen Collection and Handling

For clinical samples follow appropriate techniques for handling specimens as per established guidelines (3,4).

For food and dairy samples, follow appropriate techniques for sample collection and processing as per guidelines (5,6).

For water samples, follow appropriate techniques for sample collection, processing as per guidelines and local standards.(1)

After use, contaminated materials must be sterilized by autoclaving before discarding.

### Warning and Precautions :

In Vitro diagnostic Use only. Read the label before opening the container. Wear protective gloves/protective clothing/eye protection/ face protection. Follow good microbiological lab practices while handling specimens and culture. Standard precautions as per established guidelines should be followed while handling clinical specimens. Safety guidelines may be referred in individual safety data sheets

## Limitations :

1. Besides *Enterobacteriaceae*, other gram negative bacteria and yeasts may also grow.
2. Avoid exposure of the medium to light, as it may lead to photo oxidation and decrease productivity of the medium.
3. Overheating of the medium must be avoided, as it may destroy the productivity of the medium.
4. Further biochemical tests must be carried out for further confirmation.

## Performance and Evaluation

Performance of the medium is expected when used as per the direction on the label within the expiry period when stored at recommended temperature.

## Quality Control

### Appearance

Light pink to purple homogeneous free flowing powder

### Gelling

Firm, comparable with 1.5% Agar gel

### Colour and Clarity of prepared medium

Orangish pink coloured, clear to slightly opalescent gel with fine precipitate forms in Petri plates.

### Reaction

Reaction of 4.15% w/v aqueous solution at 25°C. pH : 7.5±0.2

### pH

7.30-7.70

### Cultural Response

M029: Cultural characteristics observed after an incubation at 35-37°C for 18-24 hours.

| Organism   | Inoculum (CFU)   | Growth         | Recovery | Colour of Colony                     |
|--|------------------|----------------|----------|--------------------------------------|
| <b>Cultural Response</b>                                       |                  |                |          |                                      |
| <i>Bacillus subtilis subsp. spizizenii</i> ATCC 6633 (00003*)  | ≥10 <sup>3</sup> | inhibited      | 0%       |                                      |
| * <i>Klebsiella aerogenes</i> ATCC 29212 (00087*)              | 50-100           | good-luxuriant | ≥50%     | pink                                 |
| <i>Enterococcus faecalis</i> ATCC 29212 (00087*)               | 50-100           | none-poor      | ≤10%     | pink, small                          |
| <i>Escherichia coli</i> ATCC 25922 (00013*)                    | 50-100           | good-luxuriant | ≥50%     | pink to rose red with metallic sheen |
| <i>Klebsiella pneumoniae</i> ATCC 13883 (00097*)               | 50-100           | good-luxuriant | ≥50%     | pink, mucoid                         |
| <i>Proteus vulgaris</i> ATCC 13315                             | 50-100           | good-luxuriant | ≥50%     | colourless to pale pink              |
| <i>Pseudomonas aeruginosa</i> ATCC 27853 (00025*)              | 50-100           | good-luxuriant | ≥50%     | colourless, irregular                |
| <i>Salmonella Typhi</i> ATCC 6539                              | 50-100           | good-luxuriant | ≥50%     | colourless to pale pink              |
| <i>Staphylococcus aureus subsp. aureus</i> ATCC 25923 (00034*) | ≥10 <sup>3</sup> | inhibited      | 0%       |                                      |
| <i>Enterobacter cloacae</i> ATCC 13047 (00083*)                | 50-100           | good           | 40-50%   | pink                                 |
| <i>Salmonella Typhimurium</i> ATCC 14028 (00031*)              | 50-100           | good-luxuriant | ≥50%     | colourless                           |
| <i>Salmonella Enteritidis</i> ATCC 13076 (00030*)              | 50-100           | good-luxuriant | ≥50%     | colourless                           |
| <i>Shigella flexneri</i> ATCC 12022 (00126*)                   | 50-100           | good-luxuriant | ≥50%     | colourless                           |

#- Formerly known as *Enterobacter aerogenes*

Key : \*Corresponding WDCM numbers.

Please refer disclaimer Overleaf.

## Storage and Shelf Life

Store between 10-30°C in a tightly closed container and the prepared medium at 2-8°C. Use before expiry date on the label. On opening, product should be properly stored dry, after tightly capping the bottle in order to prevent lump formation due to the hygroscopic nature of the product. Improper storage of the product may lead to lump formation. Store in dry ventilated area protected from extremes of temperature and sources of ignition. Seal the container tightly after use. Use before expiry date on the label.

Product performance is best if used within stated expiry period.

## Disposal

User must ensure safe disposal by autoclaving and/or incineration of used or unusable preparations of this product. Follow established laboratory procedures in disposing of infectious materials and material that comes into contact with clinical sample must be decontaminated and disposed of in accordance with current laboratory techniques (3,4).

## Reference

1. Baird R.B., Eaton A.D., and Rice E.W., (Eds.), 2015, Standard Methods for the Examination of Water and Wastewater, 23rd ed., APHA, Washington, D.C.
2. Endo S., 1904, Zentralbl. Bakteriologie, Abt. 1, Orig.35:109-11
3. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2<sup>nd</sup> Edition.
4. Jorgensen, J.H., Pfaller, M.A., Carroll, K.C., Funke, G., Landry, M.L., Richter, S.S and Warnock, D.W. (2015) Manual of Clinical Microbiology, 11th Edition. Vol. 1.
5. Salfinger Y., and Tortorello M.L. Fifth (Ed.), 2015, Compendium of Methods for the Microbiological Examination of Foods, 5th Ed., American Public Health Association, Washington, D.C.
6. Wehr H. M. and Frank J. H., 2004, Standard Methods for the Microbiological Examination of Dairy Products, 17th Ed., APHA Inc., Washington, D.C.

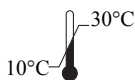
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In vitro diagnostic medical device



CE Marking



Storage temperature



Do not use if package is damaged



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