



## Deoxycholate Citrate Agar

M065

### Intended Use:

Recommended for the isolation of enteric pathogens particularly *Salmonella* and *Shigella* species.

### Composition\*\*

Ingredients	Gms / Litre
HI solids #	10.000
Proteose peptone	10.000
Lactose	10.000
Sodium deoxycholate	5.000
Neutral red	0.020
Sodium citrate	20.000
Ferric ammonium citrate	2.000
Agar	13.500
Final pH ( at 25°C)	7.5±0.2

\*\*Formula adjusted, standardized to suit performance parameters

# Equivalent to Heart Infusion solids

### Directions

Suspend 70.52 grams in 1000 ml of distilled water. Heat to boiling to dissolve the medium completely. DO NOT AUTOCLAVE. Avoid excessive heating as it is detrimental to the medium. Cool to 45-50°C. Mix well and pour into sterile petri plates.

### Principle And Interpretation

Deoxycholate Citrate Agar is prepared as per the modified formula of Leifson (4). This medium is used for the isolation and maximum recovery of intestinal pathogens belonging to *Salmonella* and *Shigella* groups from foods (5). However, it is recommended to use less inhibitory medium when *Shigellae* have to be isolated (1). The selectivity of this medium permits the use of fairly heavy inocula without danger of overgrowth of *Shigella* and *Salmonella* by other microflora. For the routine examination of stool and urine specimens, it is suggested that other media such as MacConkey Agar (M082), Bismuth Sulphite Agar (M027) etc. be used in conjunction with this medium.

This medium is similar to deoxycholate agar in comparison but is moderately more selective for enteric pathogens owing to increased concentrations of both citrate and deoxycholate salts. Sodium deoxycholate at pH 7.3 to 7.5 is inhibitory for gram-positive bacteria. Citrate salts, in the concentration included in the formulation, are inhibitory to gram-positive bacteria and most other normal intestinal organisms.

HI solids is a source of carbon and nitrogen and this ingredient is used because the inhibition of coliforms produced is greater than when an extract or simple peptone is used. Proteose peptone provides carbon, nitrogen, vitamins and minerals. Coliform bacteria and gram-positive bacteria are inhibited or greatly suppressed due to sodium deoxycholate, sodium citrate and ferric ammonium citrate. Dipotassium phosphate buffers the medium. Lactose helps in differentiating enteric bacilli, as lactose fermenters produce red colonies while lactose non-fermenters produce colourless colonies. Coliform bacteria, if present form pink colonies on this medium. The degradation of lactose causes acidification of the medium surrounding the relevant colonies and the pH indicator neutral red changes its colour to red. These colonies usually are also surrounded by a turbid zone of precipitated deoxycholic acid due to acidification of the medium. Sodium deoxycholate combines with neutral red in an acidic environment, causing the dye to go out of the solution with the subsequent precipitation of deoxycholate (4). The reduction of ferric ammonium citrate to iron sulfide is indicated by the formation of black iron sulfide. *Salmonella* and *Shigella* species do not ferment lactose but *Salmonella* may produce H<sub>2</sub>S, forming colorless colonies with or without black centers.

Citrate and iron (Fe) combination has a strong hydrolyzing effect on agar when the medium is heated, producing a soft and unelastic agar. If autoclaved the agar becomes soft and almost impossible to streak. *Salmonella Gallinarum* is inhibited

**Please refer disclaimer Overleaf.**

if sodium deoxycholate concentration is increased to 0.1 % or greater (4). Surface colonies of non-lactose fermenters often absorb a little colour (pinkish) from the medium and organisms may be mistaken for coliforms (4).

## Type of specimen

Clinical- faeces.

## Specimen Collection and Handling

For clinical samples follow appropriate techniques for handling specimens as per established guidelines (4,5).

After use, contaminated materials must be sterilized by autoclaving before discarding.

## Warning and Precautions

In Vitro diagnostic use only. Read the label before opening the container. Wear protective gloves/protective clothing/ eye protection/face protection. Follow good microbiological lab practices while handling specimens and culture. Standard precautions as per established guidelines should be followed while handling clinical specimens. Safety guidelines may be referred in individual safety data sheets.

## Limitations

1. Further Biochemical identification is required for confirmation of species.
2. Due to nutritional variations some organisms may show poor growth.

## Performance and Evaluation

Performance of the medium is expected when used as per the direction on the label within the expiry period when stored at recommended temperature.

## Quality Control

### Appearance

Light yellow to pinkish beige homogeneous free flowing powder

### Gelling

Firm, comparable with 1.35% Agar gel.

### Colour and Clarity of prepared medium

Reddish orange coloured, clear to slightly opalescent gel forms in Petri plates

### Reaction

Reaction of 7.05% w/v aqueous solution at 25°C. pH : 7.5±0.2

### pH

7.30-7.70

### Cultural Response

Cultural characteristics observed after an incubation at 35-37°C for 18-24 hours.

Organism	Growth	Inoculum (CFU)	Recovery	Colour of Colony	H <sub>2</sub> S
<b>Cultural Response</b>					
<i>Enterococcus faecalis</i> ATCC 29212 (00087*)	inhibited	>=10 <sup>3</sup>	0%		
<i>Escherichia coli</i> ATCC 25922 (00013*)	poor	50-100	20-30%	pink with bile precipitate	negative reaction
<i>Salmonella Enteritidis</i> ATCC 13076 (00030*)	good-luxuriant	50-100	>=50%	colourless	positive reaction, black centered colonies
<i>Salmonella Typhimurium</i> ATCC 14028 (00031*)	good-luxuriant	50-100	>=50%	colourless	positive reaction, black centered colonies
<i>Shigella flexneri</i> ATCC 12022 (00126*)	good	50-100	40-50%	colourless	
<i>Escherichia coli</i> ATCC 8739 (00012*)	poor	50-100	20-30%	pink with bile precipitate	negative reaction

<i>Escherichia coli</i> NCTC 9002	poor	50-100	20-30%	pink with bile precipitate	negative reaction
<i>Salmonella Abony</i> NCTC 6017 (00029*)	good-luxuriant	50-100	>=50%	colourless	positive reaction, black centered colonies
<i>Staphylococcus aureus</i> subsp. aureus ATCC 25923 (00034*)	inhibited	>=10 <sup>3</sup>	0%		

Key : \*Corresponding WDCM numbers.

## Storage and Shelf Life

Store below 30°C in a tightly closed container and the prepared medium at 20-30°C. Use before expiry date on the label. On opening, product should be properly stored dry, after tightly capping the bottle in order to prevent lump formation due to the hygroscopic nature of the product. Improper storage of the product may lead to lump formation. Store in dry ventilated area protected from extremes of temperature and sources of ignition Seal the container tightly after use. Use before expiry date on the label. Product performance is best if used within stated expiry period.

## Disposal

User must ensure safe disposal by autoclaving and/or incineration of used or unusable preparations of this product. Follow established laboratory procedures in disposing of infectious materials and material that comes into contact with clinical sample must be decontaminated and disposed of in accordance with current laboratory techniques (2,3).

## Reference

1. Frieker C.R., 1987, J. Appl. Bact., 63:99.
2. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2<sup>nd</sup> Edition.
3. Jorgensen, J.H., Pfaller, M.A., Carroll, K.C., Funke, G., Landry, M.L., Richter, S.S and Warnock., D.W. (2015) Manual of Clinical Microbiology, 11th Edition. Vol. 1.
4. Leifson, 1935, J. Path. Bact., 40:581.
5. Salfinger Y., and Tortorello M.L. Fifth (Ed.), 2015, Compendium of Methods for the Microbiological Examination of Foods, 5th Ed., American Public Health Association, Washington, D.C.

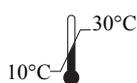
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In vitro diagnostic medical device



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