



M-Lauryl Sulphate Broth

M1023

M-Lauryl Sulphate Broth is used for enumeration of *Escherichia coli* and coliforms in water, using membrane filter technique.

Composition**

Ingredients	Gms / Litre
Peptic digest of animal tissue	39.000
Yeast extract	6.000
Lactose	30.000
Phenol red	0.200
Sodium lauryl sulphate	1.000
Final pH (at 25°C)	7.4±0.2

**Formula adjusted, standardized to suit performance parameters

Directions

Suspend 76.2 grams in 1000 ml distilled water. Heat if necessary to dissolve the medium completely. Dispense as desired and sterilize by steaming for 30 minutes on three consecutive days or by autoclaving at 15 lbs pressure (121°C) for 15 minutes.

Principle And Interpretation

The membrane filter technique is used to test relatively large volumes of samples. It is extremely useful in monitoring drinking water and a variety of natural waters (1). The earlier medium used to detect coliforms in water employed bile salts as the selective agent. This was replaced with Teepol by Burman (2). The effectiveness of teepol was demonstrated earlier (3, 4). M-Lauryl Sulphate Broth is similar to this medium, the only difference being the use of sodium lauryl sulphate as the inhibitory agent instead of teepol. M-Lauryl Sulphate Broth is recommended for enumeration of *Escherichia coli* and coliforms using membrane filtration technique (5, 6).

An absorbent pad is saturated with M-Lauryl Sulphate Broth. The filter, through which the water sample is passed, is aseptically placed on this saturated absorbent pad, with face upwards.

Burman (7) recommended the following incubation temperatures and durations.

Unchlorinated waters:

Coliform organisms : 4 hours at 30°C followed by 14 hours at 35°C

Escherichia coli : 4 hours at 30°C followed by 14 hours at 44°C

Non-chlorinated organisms benefit from 4 hours incubation at 30°C but chlorinated organisms require 6 hours incubation at 25°C. After incubation, yellow colonies are formed which should be confirmed further.

Peptic digest of animal tissue and yeast extract act as a source of nitrogen, carbon and amino acids. Lactose is the source of fermentable carbohydrate. Phenol red serves as an indicator. Sodium lauryl sulphate inhibits gram-positive bacteria.

Quality Control

Appearance

Light yellow to pink homogeneous free flowing powder

Colour and Clarity of prepared medium

Red coloured clear solution without any precipitate

Reaction

Reaction of 7.62% w/v aqueous solution at 25°C. pH : 7.4±0.2

pH

7.20-7.60

Cultural Response

M1023: Cultural characteristics on membrane filter after an incubation at different temperatures for 24 hours

Organism	Inoculum (CFU)	Growth at 35-37°C	Growth at 44°C	Colour of Colony on Membrane
<i>Enterobacter aerogenes</i> ATCC 13048	50-100	luxuriant	inhibited	yellow
<i>Bacillus subtilis</i> ATCC 6633	$\geq 10^3$	inhibited	inhibited	
<i>Staphylococcus aureus</i> ATCC 25923	$\geq 10^3$	inhibited	inhibited	
<i>Enterococcus faecalis</i> ATCC 29212	$\geq 10^3$	inhibited	inhibited	
<i>Escherichia coli</i> ATCC 25922	50-100	luxuriant	luxuriant	yellow

Storage and Shelf Life

Store below 30°C in tightly closed container and the prepared medium at 2 - 8°C. Use before expiry date on the label.

Reference

1. Eaton A. D., Clesceri L. S. and Greenberg A. E., (Eds.), 1995, Standard Methods for the Examination of Water and Wastewater, 19th Ed., American Public Health Association, Washington, D.C.
2. Burman N. P., 1967, Proc. Soc. Wat. Treat. Exam., 16:40.
3. Jameson J. E. and Emberley N.W., 1956, J. Gen. Microbiol. 15:198-204
4. Jebb W. H. H., 1959, J. Hyg. Camb. 57. 184-192
5. Joint Committee of PHLS and the Standing Committee of Analysts, 1980, J. Hyg. Camb. 85.181
6. Department of the Environmental Health and Social Security and PHLS, 1982, The Bacteriological Examination of Drinking Water Supplies, Report on Public Health and Medical Subjects No. 71, HMSO, London.
7. Burman N. P., 1967, Rec. Adv. in Bacteriological Examination of waters; C.H. Collins (Ed.), Butterworth, London.

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