



Modified AEA Sporulation Medium Base

M1236

Modified AEA Sporulation Medium is used for early sporulation of *Clostridium perfringens* from foods.

Composition**

Ingredients	Gms / Litre
Biopeptone	10.000
Yeast extract	10.000
Disodium phosphate	4.360
Monopotassium phosphate	0.250
Ammonium acetate	1.500
Magnesium sulphate. heptahydrate	0.200
Final pH (at 25°C)	7.8±0.2

**Formula adjusted, standardized to suit performance parameters

Directions

Suspend 26.31 grams in 1000 ml distilled water. Heat if necessary to dissolve the medium completely. Dispense the medium in 15 ml amounts in screw capped tubes and sterilize by autoclaving at 15 lbs pressure (121°C) for 15 minutes. Cool to 45-50°C and add 0.6 ml of filter sterilized 10% raffinose and 0.2 ml each of sterile 0.66 M sodium carbonate and 0.32% cobalt chloride dropwise to each 15 ml base medium in the tubes. Just before using, steam the medium for 10 minutes and after cooling, add 0.2 ml of filter sterilized (freshly prepared) 1.5% sodium ascorbate to each tube of the medium.

Principle And Interpretation

Clostridium perfringens causes food poisoning in humans. Foods may become contaminated with *C. perfringens* at the abattoir, in transit to shops and market places, etc. The organism is present as vegetative cells in foods (1). A heat labile enterotoxin produced only by sporulating cells induces the major symptoms of diarrhea in perfringens poisoning. Although the enterotoxin is not preformed in the food, the foods in which conditions are favorable for sporulation may contain the toxin. To determine the enterotoxigenicity of *C. perfringens* from food or faeces, it is necessary to induce sporulation in the organisms. Modified AEA Sporulation Medium (2) recommended by APHA (3), is a modification of the original medium of Taniguti (4) and is recommended for the early sporulation of *C. perfringens* from foods.

Biopeptone and yeast extract serve as essential sources of nutrients required by bacterial metabolism. Disodium phosphate buffers the medium well. Ammonium acetate, cobalt chloride, sodium carbonate and magnesium sulphate serve as sources of ions required for sporulation. Raffinose is the fermentable carbohydrate. *C. perfringens* ferments raffinose to produce acid. To test for acid, transfer 1 ml of culture to a test tube or spot plate and add 2 drops of 0.04% bromothymol blue. A yellow colour indicates acid production.

Inoculate about 2 grams of sample in Chopped Liver Broth (M606). Tubes showing turbidity after an incubation at 35-37°C for 20-24 hours are inoculated onto Perfringens Agar Base (M837). Presumptive *C. perfringens* are further confirmed by performing biochemical tests. Subculture isolates to be tested for enterotoxin in Cooked Meat Medium (M149) and incubate at 35-37°C for 24-48 hours. After incubation, transfer 2-3 drops into Fluid Thioglycollate Medium (M009) and incubate at 35-37°C for 18-24 hours. Re-incubate fresh Fluid Thioglycollate Medium (M009) with this 24 hours old culture. Incubate at 35-37°C for 4 hours. Use the 4 hours subculture to inoculate 15 ml of Modified AEA Sporulation Medium (M1236) and incubate at 35-37°C for 18-24 hours. Check spore formation by examining stained smears (3).

Quality Control

Appearance

Cream to yellow homogeneous free flowing powder

Colour and Clarity of prepared medium

Yellow coloured clear to slightly opalescent solution

Reaction

Reaction of 2.63% w/v aqueous solution at 25°C. pH : 7.8±0.2

pH

7.60-8.00

Cultural Response

M1236: Cultural characteristics observed after an incubation at 35-37°C for 48-72 hours with added sterile 10% raffinose, sodium carbonate and cobalt chloride solution and sodium ascorbate solution.

Organism	Inoculum (CFU)	Growth	Raffinose fermentation	Sporulation (observed by examining stained slides)
<i>Clostridium perfringens</i> ATCC 12924	50-100	good-luxuriant	positive reaction	positive
<i>Clostridium sporogenes</i> ATCC 11437	50-100	good-luxuriant	negative reaction	positive

Storage and Shelf Life

Store below 30°C in tightly closed container and the prepared medium at 2 - 8°C. Use before expirydate on the label.

Reference

1. Collee J. G., Fraser A. G., Marmion B. P., Simmons A., (Eds.), Mackie and McCartney, Practical Medical Microbiology, 1996, 14th Edition, Churchill Livingstone.
2. Harmon S. M., and Kautter D. A., 1986, J. Food Prot. 49:706.
3. Downes F. P. and Ito K., (Eds.), 2001, Compendium of Methods for the Microbiological Examination of Foods, 4th Ed., APHA, Washington, D.C.
4. Taniguti T., 1969, J. Food Hyg. Soc. Jpn. 9: 219.

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