



## Ascospore Agar

M804

Ascospore Agar is recommended for enrichment and detection of ascosporegenous yeasts.

### Composition\*\*

Ingredients	Gms / Litre
Yeast extract	2.500
Dextrose	1.000
Potassium acetate	10.000
Agar	30.000
Final pH ( at 25°C)	6.4±0.2

\*\*Formula adjusted, standardized to suit performance parameters

### Directions

Suspend 43.50 grams in 1000 ml distilled water. Heat to boiling to dissolve the medium completely. Sterilize by autoclaving at 15 lbs pressure (121°C) for 15 minutes. Mix well and pour into sterile Petri plates.

### Principle And Interpretation

Ascospore Agar is recommended for the enrichment and detection of ascospores in ascosporegenous yeasts such as *Saccharomyces cerevisiae*. It is based on the formula developed by McClary et al (1). Ascospore Agar is the modification of McClary medium with the addition of potassium acetate in place of sodium acetate. Acetate salt of potassium was found to be superior to sodium salt for sporulation in *Saccharomyces* (1, 2).

Dextrose and yeast extract provide the nutrients required for the growth and also stimulate ascospore formation in yeasts. Slightly acidic pH of the medium favours the growth of *Saccharomyces cerevisiae*.

### Quality Control

#### Appearance

Light yellow to brownish yellow homogeneous free flowing powder

#### Gelling

Firm, comparable with 3.0% Agar gel

#### Colour and Clarity of prepared medium

Medium amber coloured clear to slightly opalescent gel forms in Petri plates

#### Reaction

Reaction of 4.35% w/v aqueous solution at 25°C. pH : 6.4±0.2

#### pH

6.20-6.60

#### Cultural Response

M804: Cultural characteristics observed after an incubation at 25-30°C for upto 3-6 days .

Organism	Inoculum (CFU)	Growth	Recovery	Ascospores
<i>Candida albicans</i> ATCC 10231	50-100	luxuriant	≥50%	negative
<i>Saccharomyces cerevisiae</i> ATCC 9763	50-100	luxuriant	≥50%	positive

### Storage and Shelf Life

Store below 30°C in tightly closed container and the prepared medium at 2 - 8°C. Use before expiry date on the label.

### Reference

1. McClary D.O., Nulty W.L. and Miller G.R., 1959, J.Bacteriol., 78:362

2.MacFaddin J.F.,1985, Media for Isolation-Cultivation-Identification-Maintenance of Medical Bacteria, Williams and Wilkins, Baltimore.

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