

Technical Data

Tergitol-7 Agar H

M850

Tergitol-7 Agar H is recommended for selective isolation and differentiation of enteric bacteria from urine specimens.

Composition**				
Ingredients	Gms / Litre			
Proteose peptone	5.000			
Yeast extract	3.000			
Lactose	10.000			
Ferric ammonium citrate	0.500			
Sodium thiosulphate	0.500			
Bromo thymol blue	0.025			
Sodium heptadecyl sulphate(Tergitol-7)	0.100			
Agar	15.000			
Final pH (at 25°C)	7.2±0.2			
**Formula adjusted, standardized to suit performance parameters				

Directions

Suspend 34.13 grams in 1000 ml distilled water. Heat to boiling to dissolve the medium completely. Sterilize by autoclaving at 15 lbs pressure (121°C) for 15 minutes. Cool to 45-50°C. Aseptically add 3 ml of 1% 2,3,5 Triphenyl Tetrazolium Chloride (TTC) Solution (FD057), if desired. Mix well and pour into sterile Petri plates.

Principle And Interpretation

Tergitol-7 Agar was originally designed by Chapman (1) and later on modified by incorporating 2,3,5-Triphenyl Tetrazolium Chloride (TTC) into the medium. This medium is selective and differential used for the detection and enumeration of coliform organisms. Pollard (2) has reported the selective bactericidal property of sodium heptadecyl sulphate (Tergitol-7). Kulp et al (3) corroborated the use of Tergitol-7 Agar with TTC in routine analysis of water and Mossel (4) used this medium for the examination of food materials.

Tergitol-7 Agar H, is a modification of Chapman formulation (1) used for selective isolation and differentiation of enteric bacilli from urine samples. It contains sodium thiosulphate as an indicator of H2S production. H2S producing bacteria form black colonies or colonies with black centres.

Proteose peptone and yeast extract serve as sources of carbon, nitrogen and other essential nutrients including vitamin B complex. Sodium heptadecyl sulphate (Tergitol-7) inhibits gram-positive bacteria and *Proteus* swarming and yields better recovery of coliforms. Bromo thymol blue is the pH indicator. Lactose fermenting organisms form yellow colonies with yellow zones while *Klebsiella* and *Enterobacter* form greenish yellow colonies. Lactose non-fermenters produce blue colonies. TTC is reduced in the bacterial cell to form formazan, a red coloured insoluble complex, thereby producing red coloured colonies.

Quality Control

Appearance

Cream to light green homogeneous free flowing powder

Gelling Firm, comparable with 1.5% Agar gel Colour and Clarity of prepared medium Green coloured clear to slightly opalescent gel forms in Petri plates Reaction

Reaction of 3.41% w/v aqueous solution at 25°C. pH : 7.2±0.2

pH 7.00-7.40

Please refer disclaimer Overleaf.

Cultural Response

M850: Cultural characteristics observed after an incubation at 35-37°C for 18-48 hours, with added TTC solution 1% (FD057)

Organism	Inoculum (CFU)	Growth	Recovery	Colour of colony	H2S
Escherichia coli ATCC 25922	50-100	good-luxuriant	>=50%	yellow	negative
Proteus mirabilis ATCC 25933	50-100	good-luxuriant	>=50%	blue	positive
Klebsiella pneumoniae ATCC 13883	50-100	fair-good	30-40%	greenish yellov	vnegative
Salmonella Enteritidis ATC 13076	C50-100	good-luxuriant	>=50%	blue	positive
Enterococcus faecalis ATCO 29212	C>=10 ³	inhibited	0%		

Storage and Shelf Life

Store below 30°C in tightly closed container and the prepared medium at 2 - 8°C. Use before expiry date on the label.

Reference

- 1. Chapman G.H., 1947, J. Bact., 53:504.
- 2. Pollard A.L., 1946, Science, 103:758.
- 3. Kulp W., Mascoli C. and Tavshanjian O., 1953, Am. J. Public Health, 43:1111.
- 4. Mossel D.A.A., 1962, J. Appl. Bact., 25:20.

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HiMedia Laboratories Pvt. Ltd. A-516, Swastik Disha Business Park, Via Vadhani Ind. Est., LBS Marg, Mumbai-400086, India. Customer care No.: 022-6147 1919 Email: techhelp@himedialabs.com

