

# **Technical Data**

# **MUG EC 0157 Agar**

M1373

MUG EC O157 Agar is recommended for isolation and differentiation of enterohaemorrhagic *Escherichia coli* O157:H7 from foodstuffs, water and clinical samples by a fluorogenic method.

# Composition\*\*

*	
Ingredients	Gms / Litre
Casein peptone	20.000
Meat extract	2.000
Yeast extract	1.000
Sorbitol	10.000
Ferric ammonium citrate	0.500
Sodium chloride	5.000
Bromothymol blue	0.025
Sodium thiosulphate	2.000
Sodium deoxycholate	1.120
4-Methylumbelliferyl β-D-Glucuronide (MUG)	0.100
Agar	13.000
Final pH (at 25°C)	$7.4\pm0.2$

<sup>\*\*</sup>Formula adjusted, standardized to suit performance parameters

#### **Directions**

Suspend 54.74 grams in 1000 ml distilled water. Heat to boiling to dissolve the medium completely. Sterilize by autoclaving at 15 lbs pressure (121°C) for 15 minutes and pour into sterile Petri plates.

# **Principle And Interpretation**

MUG EC O157 Agar is recommended (1) for isolation and enumeration of enterohaemorrhagic *Escherichia coli* (EHEC) from foodstuffs, water and clinical samples based on sorbitol utilization and formation of beta-glucuronidase enzyme. The enterohaemorrhagic *E. coli* O157:H7 strains produce toxins, which can result in life threatening extra intestinal complications in the form of the hemolytic uremic syndrome and thrombotic-thrombocytopenic purpura. Due to severe clinical implications, the isolation and detection of *E. coli* O157:H7 strains are of importance.

Sodium deoxycholate inhibits the growth of gram-positive microbes. Sorbitol provides carbon and energy source. Bromothymol blue is the pH indicator. Microorganisms utilizing sorbitol exhibit yellow colonies whereas sorbitol-negative strains (such as *E.coli* O157:H7) grow as greenish colonies. Hydrogen suphide production is detected as black-brown colony colouration due to presence of sodium thiosulphate and ferric ammonium citrate. Thus *Proteus mirabilis* having similar biochemical characteristics as that of *E. coli* O157:H7 can easily be differentiated. 4-Methylumbelliferyl b-D-glucuronide (MUG) is converted into 4-methylumbelliferone by beta-D-glucuronidase-forming pathogens. 4-methylumbelliferone fluoresces under UV light. All commensal *E. coli* produce beta-glucuronidase. *E. coli* O157:H7 is not capable of forming b-glucuronidas, thus when exposed under long-wave UV light, no fluorescence is observed. The plates were exposed to ammonia fumes to increase fluorescence as suggested by Freir and Hartman (2).

# **Quality Control**

#### **Appearance**

Cream to yellow homogeneous free flowing powder

#### Gelling

Firm, comparable with 1.3% Agar gel.

### Colour and Clarity of prepared medium

Bluish green coloured clear to slightly opalescent gel forms in Petri plates

#### Reaction

Reaction of 5.47% w/v aqueous solution at 25°C. pH: 7.4±0.2

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## pН

7.20-7.60

#### **Cultural Response**

M1373: Cultural characteristics observed after an incubation at 35-37°C for 18-24 hours.

Organism	Inoculum (CFU)	Growth	Recovery	Colour of colony	Fluorescence (under UV)*
Cultural Response					
Enterobacter aerogenes ATCC 13048	50-100	luxuriant	>=50%	yellow	negative
Escherichia coli O157:H7	50-100	luxuriant	>=50%	colourless	negative
Escherichia coli ATCC 25922	50-100	luxuriant	>=50%	yellow	positive
Enterococcus faecalis ATCO 19433	$C >= 10^3$	inhibited	0%		
Proteus mirabilis ATCC 25933	50-100	luxuriant	>=50%	brown, may show black colouration(H production)	negative 2S
Salmonella Typhimurium ATCC 14028	50-100	luxuriant	>=50%	yellow w/black negative centre	

Key: \* - Fluorescence can be visualized on addition of NaOH solution or exposure to ammonia fumes.

# **Storage and Shelf Life**

Store below 30°C in tightly closed container and the prepared medium at 2 - 8°C. Use before expiry date on the label.

#### Reference

- 1. Szabo R. A., Todd E. C., Jean A., 1986, J. Food Prot., 10:768-772.
- 2.Freir T.A. and Hartman P.A. (1987) Appl. Env. Microbiol. 53. 1246-1250

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