

# **Technical Data**

# Gillies Agar No. 2 (Sucrose Salicin Agar)

**M240** 

Gillies Agar No. 2 (Sucrose Salicin Agar) is recommended for detection of motility, hydrogen sulphide, indole production and fermentation of sucrose and salicin for identification of *Salmonella* and *Shigella* species.

### **Composition\*\***

Ingredients	Gms / Litre
Peptic digest of animal tissue	10.000
Casein enzymic hydrolysate	10.000
Sodium chloride	5.000
Disodium phosphate	0.250
Sucrose	10.000
Salicin	10.000
Bromothymol blue	0.010
Sodium thiosulphate	0.025
Agar	3.000
Final pH ( at 25°C)	7.4±0.2

\*\*Formula adjusted, standardized to suit performance parameters

## Directions

Suspend 48.28 grams in 1000 ml distilled water. Heat to boiling to dissolve the medium completely. Distribute in tubes and sterilize by autoclaving at 118° - 121°C for 15 minutes(12-15 lbs pressure). Allow the tubes to cool in an upright position. Suspend Kovacs reagent strips and lead acetate papers from the cap or the cotton plug over the medium but not touching the surface of the medium.

# **Principle And Interpretation**

*Enterobacteriaceae* genera consist of gram-negative bacilli and are widely distributed in nature. It includes pathogens such as *Salmonella, Shigella, Yersinia*, diarrheagenic *E.coli* and others. These bacteria cause multitude of diseases in humans and are frequently isolated from clinical specimens. Detection and identification of the bacteria are of importance both from clinical and epidemiological point of view. The other enterobacteria are essentially commensals or saprophytes (1). Gillies Agar No. 2 (2), a modification of Kohns Medium (3) is recommended for detection of motility, hydrogen sulphide, indole production and fermentation of sucrose and salicin. This medium is a reliable substitute for the conventional method of determining the biochemical identity of non-lactose fermenting colonies prior to confirmation by serological typing (1).

Fermentation of sucrose and salicin leads to acid production that causes the pH indicator dye, bromothymol blue, to change from blue to yellow. The accompanying gas production during fermentation causes bubbles to form, which appears in varying degrees from a slight splitting along the wire track to disruption of the medium. Non-motile organisms grow only along the line of inoculation whereas motile species show either a diffuse even growth spreading from the inoculum or more rarely localized outgrowths, which are usually fan shaped or occasionally nodular. Hydrogen sulphide production causes blackening of the lead acetate paper and the formation of indole gives a red colour in the Kovacs reagent strips.

Peptic digest of animal tissue and casein enzymic hydrolysate serve as sources of essential nutrients for bacterial growth. Sodium chloride maintains the osmotic equilibrium of the medium. Sucrose and salicin are the fermentable carbohydrates with bromothymol blue as the pH indicator. Sodium thiosulphate aids in the production of hydrogen sulphide.

The specimen is inoculated into a preliminary enrichment medium such as Fluid Tetrathionate Broth Base (M032). After incubation at 35-37°C for 18-24 hours, this enriched culture is subcultured on a differential media such as Wilson and Blair Medium (M331) or MacConkey Agar (M081). Presumptive colonies are purified and pure cultures are used to inoculate the tubes of Gillies Agar No. 2.

Gillies Medium No. 2 is used by stab inoculating one half the depth of the medium using a straight needle. Kovacs reagent strips and lead acetate papers can be suspended from the cap or with the cotton plug over the medium but not touching the surface of the medium.

#### **Quality Control**

#### Appearance

Light yellow to light green homogeneous free flowing powder

#### Gelling

Semisolid, comparable with 0.3% Agar gel.

#### Colour and Clarity of prepared medium

Green coloured, clear to slightly opalescent gel forms in tubes as butts

#### Reaction

Reaction of 4.83% w/v aqueous solution at 25°C. pH : 7.4±0.2

#### pН

7.20-7.60

#### **Cultural Response**

M240: Cultural characteristics observed after an incubation at 35 - 37°C for 18 - 24 hours.

Organism	Inoculum (CFU)	Growth	H2S	Indole	Motility	Sucrose/ Salicin
Proteus vulgaris ATCC 13315	50-100	luxuriant	weak reaction	weak reaction	positive, growth away from stabline causing turbidity	positive reaction,yellow colouration of the medium
Salmonella Typhi ATCC 6539	50-100	luxuriant	weak reaction	negative reaction,no colour development/ cloudy ring	positive, growth away from stabline causing turbidity	negative reaction
Shigella sonnei ATCC 2593.	7 50-100	luxuriant	negative reaction	negative reaction, no colour development / cloudy ring	negative growth along the stabline, surrounding medium remains clear	negative reaction

#### **Storage and Shelf Life**

Store below 30°C in tightly closed container and prepared medium at 2-8°C. Use before expiry period on the label.

#### Reference

1.Cruickshank R., Duguid J. P. Marmion B. P., Swain R. H. A., (Eds.), 1975, Medical Microbiology, 12th Edition, Vol. II, Churchill Livingstone. 2.Gillies R. R., 1956, J. Clin. Pathol., 9, 368.

3.Kohn J., 1953, J. Clin. Path., 6, 249.

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